Gene stacking refers to the process of combining two or more genes of interest into a single plant. This method of gene stacking has brought many notable products such as “Golden Rice,” a rice variety that can provide a child with 50% of the total carotenoids needed for healthy vision. 

While the refuge strategy lessens the chance for the insect pest to overcome the gene, many regulatory agencies require a refuge of non-GM crops alongside the GM crop. Typically a refuge is about 20% of the total crop area for a mono-GT trait and 30% to 50% of the total crop area for a multi-GT trait.

Why Gene Stacking?

Gene stacking is especially useful in metabolic engineering of plants since most metabolic processes and biochemical pathways involve numerous genes interacting with each other. For example, this is done by combining the glyphosate resistance gene epsps with the pat gene conferring resistance to herbicide glufosinate and/or with the bcs gene conferring resistance to herbicide dicamba.

Gene stacking is also used in weed management. Weed resistance to commercial herbicides has been documented for different herbicidal modes of action. To catch up, weeds need to adapt to defeat the gene technology. Experience has shown that the resistance and durability will be greater for multiple insect resistance based on different genes than on a single one. Gene stacking allows a lower refuge area requirement that somehow limits yield.

Insect resistance management for Bt crops with the same Bt gene technology. Experience has shown that the resistance and durability will be greater for multiple insect resistance based on different genes than on a single one. Gene stacking allows a lower refuge area requirement that somehow limits yield.

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Gene Stacking Process

Table 1 summarizes the common methods of gene stacking and provides examples of commercialized stacks produced by each method. The easiest and quickest way to stack up genes into a plant is to make crosses between parental plants that have different biotech traits, an approach known as hybrid stacking. Most of the commercially available biotech stacks, like triple stack, and quadruple stack, are products of serial hybrid stacking which is widely adapted and accepted. Another method of gene stacking is known as molecular stacking which involves the introduction of gene constructs simultaneously or sequentially into the target plant by standard delivery systems such as Agrobacterium-mediated and biolistic methods. In some stacks, molecular stacking has been done with conventional breeding approaches to put together the desired traits.

**Trends in Biotech Stacks**

It is estimated that a total of 80.5 million ha were planted to biotech stacks in 2018. This accounts for more than 42 percent of the 191.7 million ha of biotech crops planted worldwide. The US Department of Agriculture estimated that 89 percent of cotton acres and 80 percent of corn acres were planted with stacked seeds in 2019.

The first stack that gained regulatory approval in 1995 was a dual hybrid cotton stack produced by crossing Bollgard™ cotton that expresses the Bt toxin cry1Ab and Roundup Ready™ cotton that produces the epsps enzyme conferring resistance to herbicide glyphosate. Following commercial success of this hybrid stack, developers sought to stack up more biotech traits into their crop portfolio to create multi-stack hybrids. The cotton triple stack which combines two Bt genes with the glyphosate resistance gene occupied more than 20 percent of the US cotton area in 2008. The recently released eight-gene maize stack known by its trade name SmartStax™ is the result of crossing four different biotech maize lines to combine two herbicide tolerance genes with six Bt genes. The resulting stack features dual modes of control for weeds, lepidopteran insects and coleopteran insects and allowed for the refuge requirement to be reduced from 20 percent to 5 percent in the US Corn Belt.

The increasing number of biotech traits in recent stacks has set the trend for the next generation of biotech crops. Future stacks are likely to involve not only multiple pest resistances but the combination of these traits with engineered metabolic pathways and simultaneous introductions of multiple pathways through metabolic engineering (for example, pathways for beta carotene, ascorbate, folate and vitamin E synthesis).3,7,8

**Regulatory Approaches**

Regulatory principles and procedures for approval and release of biotech stacks differ globally. In countries like the USA and Canada, no separate or additional regulatory approval is necessary for commercializing hybrid stacks that are products of crossing a number of already approved biotech lines. This policy is based on the argument that interactions between individual trait components in a stack that have been shown to pose no environmental or health hazard would not result in new or altered hazards. The US Environmental Protection Agency, however, may require separate safety review of a stack upon identification of a specific hazard associated with combined “plant incorporated protectants” or PIPs (eg. Bt insecticidal proteins), since combinations of PIPs may result in higher or altered toxicity.

In Japan and European Union (EU) countries to the contrary, stacks are considered new events, even if individual events have market approval, and must pass through a separate regulatory approval process, including risk assessment of their safety, similar to mono-trait biotech events. Risk assessment is focused on the identification of additional risks that could arise from the combined genes. Possible risks are altered effects of interacting proteins on the target and non-target organisms and increased invasiveness of the crop that may pose environmental risks.

**Table 1. Common gene stacking methods used in the production of biotech stacks.**

<table>
<thead>
<tr>
<th>Gene stacking method</th>
<th>Description</th>
<th>Examples of commercial stacks*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Hybrid stacking</strong></td>
<td>A plant harboring one or more transgenes is cross-hybridized with another plant containing other transgenes. Development of a multi-stack hybrid occurs via iterative hybridization.</td>
<td><strong>Maize:</strong> Agrisure™ Viptera™ 3220 (Bt11 x MIR162 x TC1507 x GA21) <strong>Cotton:</strong> Roundup Ready™ Flex Bollgard™ II (MON8913 x MON15985)</td>
</tr>
<tr>
<td><strong>Co-transformation</strong></td>
<td>A plant is transformed with two or more independent transgenes. The transgenes of interest are in separate gene constructs and delivered to the plant simultaneously.</td>
<td><strong>Maize:</strong> NaturGard™ Knockout™ (B176), BT Xtra™ (DBT418), YieldGard™ (MON810, MON809, MON802)</td>
</tr>
<tr>
<td><strong>Linked genes or multigene cassette transformation</strong></td>
<td>A plant is transformed with a single gene construct that harbors two or more linked transgenes.</td>
<td><strong>Maize:</strong> Herculex™ I (TC1507), Herculex™ RW (59122), Agrisure™ CB/LL (B11) <strong>Soybean:</strong> Vistive™ Gold (MON87705)</td>
</tr>
<tr>
<td><strong>Re-transformation</strong></td>
<td>A plant harboring a transgene is transformed with other transgenes.</td>
<td><strong>Cotton:</strong> Bollgard™ II (MON15985)</td>
</tr>
</tbody>
</table>

*The examples of commercial biotech stacks are taken from the online ISAAA GM Approval Database. Links to more detailed information on the derivation of the biotech stacks are provided therein.

**Technological Challenges**

For the developer, the choice between a large molecular stack or a complex hybrid stack will be based on the monetary cost and timeline for developing and registering a stack. In countries where a stack must pass a separate regulatory review, the one-shot molecular stacking may be more cost effective than the lengthy hybrid stacking. There are, however, technological concerns in molecular stacking which include the design of large multi-gene constructs, method of delivery into plant cells and the stability of expression of multiple genes. Molecular biologists are developing new genetic engineering approaches to address these concerns. Among the promising technologies include site-specific gene recombination systems in conjunction with the use of engineered DNA cutting enzymes and the artificial gene assembly known as minichromosome.

How the multiple transgenes might affect the overall physiology of the plant and how many genes and what combinations of genes can be stacked into a plant are of primary concern to developers. If multiple transgene insertions will alter protein synthesis and metabolic processes of the plant drastically, these likely will compromise the yield. While this yield drag is not necessarily a biosafety concern, the yield loss will need to be offset by reduced production cost and the extra premium farmers will make from the added value of the biotech trait(s) in order for a stack to be a viable and beneficial biotech product.